

Application of infrared and nuclear magnetic resonance spectra in studying the bacterial efficacy of some oxazepane derivatives derived from hydrazones

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Abstract: This research included the utilisation of infrared and nuclear magnetic resonance spectra in studying and preparing new derivatives of the oxazepan ring by a series of reactions, where the triazole ring was prepared from the reaction of the carboxylic acid with carbothiohydrazide, and from the resulting reaction (triazole) with aqueous hydrazine. The latter was reacted with benzaldehyde compensators to prepare hydrazones, which is the basic intermediate for the preparation of the oxazepan ring. The hydrazones were reacted with succinic anhydride, and the validity of the structures was proven using infrared and nuclear magnetic resonance spectra of proton and carbon. Its biological activity has also been tested for two species of Gram-positive bacteria, *Staphylococcus epidermidis* and Gram-negative *Klebsiella pneumoniae*.

Keywords: Infrared spectra; triazole; hydrazide; hydrazone; oxazepane; biological activity.

1. Introduction

The Heterocyclic compound consists of nitrogen, oxygen, and sulphate atoms in addition to carbon atoms [1]. Heterocyclic compounds are widely distributed in nature, as they enter into the composition of many organic compounds necessary for the basic formation of life, and they are also found in different forms in sugars and their derivatives [2]. Triazole, a heterocyclic organic compound containing nitrogen currently receiving attention due to its biological, industrial, and medical importance, is 2,14-triazole, as triazole is a five-membered ring containing three nitrogen atoms and two nitrogen atoms. Carbon atoms can contain two types of bonds, as the nucleus of 1,3,4-triazole has antimicrobial effects and treats hypoglycemia and hypertension [3]. Where the number (1) gives the nitrogen atom to which the hydrogen atom is attached to distinguish it. Triazole compounds, in which the nitrogen atoms are not linked to each other because they are separated by two carbon atoms, are named as follows: 1,3,4-triazole. It also gives the nitrogen atom to which it is attached. The number of hydrogen atoms (1), and the two carbon atoms (5, 2) with substituents [4].

Hydrazides and their derivatives are important intermediates used in the preparation of many heterocyclic organic compounds, as they are intermediates in the preparation of many



compounds, such as Schiff bases, due to their biological activity [5]. Hydrazones are organic compounds resulting from the condensation reaction between the hydrazine compound and various aldehydes or ketones, which occurs through an unstable intermediate state to a product known as hydrazone, which consists of 2 nitrogen atoms; the first is hydrogen, and the second is the hydrazone. The nitrogen atom. The second is linked to the carbon atom by bonding the hydrazone carrying the functional group (-HC=N-NH-C=O) [6]. Hydrazones and their derivatives have recently received great attention after the discovery of the biological effects of this compound. They are extensively applied in medical and pharmaceutical fields and have also been used as antifungals [7] and antibacterials [8]. Oxazepane contains seven atoms consisting of 5 carbon atoms, one nitrogen atom, and one oxygen atom (90), of which 1,3- Oxazepane -7,4-dione can be manufactured by adding anhydrides such as phthalic acid or maleic acid and others. A double bond of Schiff base or isomethene (C=N) of hydrazine[9]. Oxazepane compounds also contain three isomers numbered according to the two nitrogen atoms in the ring, where the nitrogen atom is in position (2, 3, or 4) and the oxygen atom is in position (1) [10].

Oxazepane compounds have wide biological importance and have received wide attention in the medical field as they have shown antibacterial [11], anticonvulsant [12], and antioxidant [13].

2. Materials and Methods

2.1. Used Chemicals: All used chemicals supplied by Aldrich, BDH Thomas, Fluka, and Merck were used.

2.2. Instruments used: The melting point was ascertained using a Shimadzu-8400S FT-IR spectrometer, a 400-4000 cm⁻¹ sulfur bromide disk, a 9300 thermometer, and 400 MHz Bruker ¹H- and ¹³C-NMR spectra. The catalyst surface was verified by SEM examination. Fluka silica gel plates 0.02cm in thickness were used for thin-layer chromatography (TLC).

2.3. Preparation of Triazole (MH1)

Equal moles (0.003 mol) of carboxylic acid were mixed with hydrazine carbothiohydrazide in a heat-resistant vessel without solvent, stirring for 10 minutes until the colour changed. The resulting compound was collected and recrystallised from ethanol, giving a yellow colour with a product yield of 83% M.p 149-151 [14].

2.4. Preparation of Hydrazide (MH2)

Equal moles (0.002 mol) of triazole MH1 were mixed with 98% aqueous hydrazine in a vessel; 20 ml of CS2 was added, stirring for 10 minutes. The resulting material was collected and recrystallised from ethanol, giving an orange colour with a yield of 79% M.p 176-178 [15].

2.5. Preparation of Hydrazones (MH3-MH7)

The resulting hydrazide MH2 was mixed with the benzaldehyde substitutes in a vessel in equal molar ratios (0.003 mol) without solvent, stirring for 8 minutes [16]. The resulting material was then collected and recrystallised from ethanol, as in Table 1.



Compound	R	Molecular formulas	m.p. °C	Yield %	Colour
MH3	4-Br	$C_{18}H_{14}BrN_7S_2$	197-199	68	Yellow
MH4	4-NO2	$C_{18}H_{14}N_8S_2O_2$	188-190	64	Brown
MH5	4-C1	$C_{18}H_{14}CIN_7S_2$	203-205	71	Whit
MH6	4-OH	$C_{18}H_{15}N_7S_2O$	200-202	63	Dark Yellow
MH7	4-H	$C_{18}H_{15}N_7S_2\\$	191-193	60	Blue

Table (1): Some physical characteristics of the synthesised compound (MH3-MH7).

2.6. Preparation of Oxazepane (MH8-MH12)

In a heat-resistant vessel, equal moles (0.006 mol) of the prepared hydrazones (MH3-MH7) were mixed with succinic anhydride without solvent, stirring for 15 min [17]. The material was then collected and recrystallised from ethanol, as in Table 2.

Compound	R	Molecular formulas	m.p. ⁰C	Yield %	Colour
MH8	4-Br	$C_{22}H_{18}BrN_7O_3S_2$	221-223	64	Light Yellow
МН9	4-NO2	$C_{22}H_{18}N_8O_5S_2$	234-236	62	Dark Brown
MH10	4-C1	$C_{22}H_{18}CIN_7O_3S_2$	237-239	68	Yellow
MH11	4-OH	$C_{22}H_{19}N_7O_4S_2$	243-245	73	Yellow
MH12	4-H	$C_{22}H_{19}N_7O_3S_2$	226-228	71	whit

Table (2): Some physical characteristics of the synthesised compound (MH8-MH12).

2.7. Study of biological activity

Two colonies of pure bacterial isolates of both Gram-positive **Staphylococcus** *epidermidis* and Gram-negative *Klebsiella pneumoniae* were transferred from the solid culture medium to test tubes containing (5 ml) distilled water using heat-sterilised holders [18,19]. The tubes were incubated at 37°C for 16-20 hrs and then diluted using a physiological solution until the turbidity reached standard turbidity levels to obtain a cell count of approximately (1.5×108) cells/ml. Chemical solutions of some of the synthesised compounds were prepared using dimethyl sulfoxide (DMSO) solvent at three concentrations (0.1, 0.01, 0.001) mg.mL⁻¹ of each substance (for each solid derivative). Agar-Miller-Hinton (MHA) medium was introduced into a test tube containing diluted bacterial growth [20,21]; the swab was pressed against the inner walls of the tube to remove the extra inoculums, followed by inoculation with a sterile swab. In order for the inoculum to be evenly distributed [22-24].



3. Results

The synthesised compounds are shown in Scheme 1.



Scheme (1): Path of the ready compounds (MH1-MH12).

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3.1. Characterisation of Triazole (MH1).

The FT-IR spectrum of MH1 showed that the absorption bands of rubber (NH₂) were present between 3431 and 3335 cm⁻¹, and the absorption bands were present at position 3191 cm⁻¹. For rubber (NH), the band at 3039 cm⁻¹ is attributed to the aromatic (=C-H) stretching, and the band at 1620 cm⁻¹ belongs to the (C=N) stretching, and the1485-1519 cm⁻¹ band is attributed to the aromatic (C=C) stretching[25], as shown in Figure 1.



Figure (1): The compound's FT-IR spectra (MH1).



During the study of the ¹H-NMR spectrum of MH1, one signal was noticed at position (5.29 ppm) attributed to the proton (NH₂), and two signals shown in the range of 6.30-6.48 ppm due to the presence of a double bond (CH=CH), several signals in the range of 7.41-7.76 ppm attributed to the protons of the aromatic ring, signals at positions (10.10) ppm belongs to the protons (NH), and a signal at position (12.22) ppm belongs to the proton (SH), then the signal of the solvent DMSO-d⁶ at position 2.50 ppm, as shown in Figure 2.



Figure (2): 1-H NMR spectra of the substance (MH1).

When the 13C-NMR spectrum of MH1 was studied, two signals were observed belonging to the double bond carbon (C=C) at the position (113.01-119.89) ppm, signals attributed to the aromatic ring carbon in the range (122.51-145.70) ppm, and two signals attributed to the (C=N) triazole ring at the position (161.34-162.13) ppm. A multiple signal in the range (39.33-40.58) ppm was attributed to the carbon of the solvent DMSO-d⁶. as shown in Figure 3.



Figure (3): ¹³C-NMR Spectra of the substance (MH1).



3.2. Characterisation of Hydrazide (MH2).

When studying the infrared spectrum of (MH2) compounds, it was found that a band is formed in the region (1616) cm⁻¹ due to (C=N) stretching, and the absorption band in the region (3344-3303) cm-1, which resulted, is often attributed to (NH₂) stretching, and a band at the site (3238,3180) cm-1 is due to (NH), and the aromatic (C-H) stretching is responsible for the absorption band in the region (3002) cm-1. The two absorption bands at (1093) cm⁻¹ are usually because of the C=S stretching, and the two bands are because of the aromatic (C=C) stretching in the range (1519-1461) [26]. As in Figure 4





The 1H NMR spectrum of MH2 showed one signal was at 4.26 ppm because of the presence of the NH₂. Two signals were present between 6.53 and 6.69 ppm because of the presence of a double bond (CH=CH), multiple signals were present in the range of 7.34-7.63 ppm because of the protons of the aromatic ring, signals at positions (7.01) ppm attributed to the proton (NH) adjacent to the triazole ring, a signal at 8.81 ppm belongs to the proton (NH) adjacent to the anine, a signal at position (9.23) ppm attributed to the proton (NH) of the five-membered ring, a signal at 10.85 ppm attributed to the proton (SH), then the signal of the solvent DMSO-d6 at position (2.50) ppm, see Figure 5.





Figure (5): 1-H NMR spectra of the substance (MH2).

When studying the 13C-NMR spectrum of MH2, two signals were observed belonging to the carbon of the double bond (C=C) at the position (112.17-113.72) ppm, two signals attributed to the carbon of the aromatic ring in the range (121.87-143.81) ppm, two signals attributed to the triazole ring (C=N) at the position (152.60-166.39) ppm, and a signal at the position (182.28) ppm due to the presence of C=C carbon. Also, multiple signals in the range (39.54-40.59) ppm due to the presence of carbon of the solvent DMSO-d6, as shown in Figure 6.



Figure (6): ¹³C-NMR Spectra of MH2.

3.3. Characterisation of Hydrazone (MH3-MH7).

When studying the infrared spectrum of hydrazones, it was found that a band is formed between 1635 and 1626 cm⁻¹ due to (C=N) stretching, and the resulting absorption bands between 3338 and 3232 cm⁻¹ are often attributed to (NH) stretching, and the aromatic (C-H) stretching band is responsible for the absorption band in the range (3060-3027) cm⁻¹. The two



absorption bands in the range (3162-3141) cm-1 are usually attributed to (=C-H) stretching, and the two bands are attributed to aromatic (C=C) stretching in the range (1553-1458) [27]. As in Table and Figures 7 and 8.

Comp.	Р	ν(C-H) Arom.	ν(=C-H)		∨(SH)	∨(C=N)	v(C=C)	Others
No.	ĸ		Olphen.		v(C=S)		Arom.	
4-Br	4-Br		3141	3240	2368	1635	1508,145 8	v(C-Br) 659
INILIO		3000		3350	1092	1589		
NAL 1.4	$4-NO_2$	2057	2454	3238	2453	1626	1545,148	v(N-O) as
MH4		3057	3151	3293	1081	1600	7	sy1514. Sy1315
	4-Cl	0007	0.100	3251	2503	1635	1517,146	
MH5		3037	3122	3338	1058	1596	0	v (C-CI)783
МЦС	4-OH	2027	2147	3246	2487	1629	1553,148	(ОЦ) 2206
		3027	3147	3305	1075	1601	1	V(UH) 3390
MH7	4- H	3034	3162	3232	2396	1631	1548,147	
		5004	5102	3287	1069	1606	9	
90 - - %T - 75 - - 60 - - 45 -	336.12	Br	2368.42-		1635.52~ 1589.23~ 1508.23~ 1458.08~	1313.43-	1091.63-	798.47
MH	300 3	00	2000	. –	1500		1000	500 1/cm

Table (3): FT-IR absorption results for Prepared compounds (MH3-MH7)







Figure (8): The compound's FT-IR spectra (MH5).

The 1H NMR spectrum of MH2 shows two signals were noticed in the range (6.71-6.83) ppm attributed to the double bond (CH=CH), and several signals in the range (7.20-7.96) ppm because of the presence of the protons of the aromatic ring, signals at positions (7.02) ppm attributed to the proton (NH) adjacent to the triazole ring, a signal at position (8.39) ppm part of the colourant because of the presence of the proton (=CH). A signal at position (9.01) ppm because of the presence of proton (NH), a signal at position (9.98) ppm attributed to the proton (NH) of the five-membered ring, a signal at position (11.47) ppm because of the presence of proton (SH), then the signal of the solvent DMSO-d6 at position (2.49) ppm million, see Figure 9.



Figure (9): 1-H NMR spectra of the substance (MH5).

When studying the 13C-NMR spectrum of MH3, two signals were observed belonging to the double bond carbon atom (C=C) at position (117.24-119.83) ppm, two signals attributed to the aromatic ring carbon atom in the range (121.72-136.34) ppm, a signal at position (141.34) ppm because of the formation of carbon (C=N), two signals attributed to the triazole ring (C=N) at position (154.00-161.13) ppm, and a signal at position (176.81) ppm attributed to the carbon



atom (C=S). A multiple signal was also observed in the range (39.30-40.56) ppm because of the presence of carbon atom of the solvent DMSO-d6, as shown in Figure 10.



Figure (10): ¹³C-NMR Spectra of the substance (MH3).

3.4. Characterisation of Oxazepane (MH8-MH12).

In the FT-IR spectrum of Oxazepane derivatives, a band was noticed in the range (1728-1703) cm^{-1} attributed to the ester (C=O) in the heptad ring, a band in the range (1670-1645) cm^{-1} attributed to the imide (C=O) in the resulting ring, a band in the range (1365-1347) cm^{-1} attributed to (C-O) in the same ring, and two bands attributed to the aliphatic (C-H) in the range (2893-2858) cm^{-1} and (2980-2928) cm^{-1} , in addition to two bands in the range (1577-1521) cm^{-1} and (1488-1462) cm^{-1} attributed to the aromatic (C=C) [28]. These are shown in Table 4 and Figures 11 and 12.

Comp. No.	R	v(C-H) Arom.	v(=C-H) Aliph.	∨(N-H)	v(SH)	∨(C=N)	v(C=C)	Others
	ĸ				v(C=S)		Arom.	
МЦО	1 Dr	2024	2876	3257	1709	1603	1522 1476	(C Pr) 617
	4-DI	3034	2943	3321	1654	1354	1525,1470	V (С-Ы) 017
мцо	4-	2015	2893	3207	1728	1612	1577 1464	v(N-O)as
	NO ₂	3043	2980	3286	1670	1365	1377,1404	Sy1323. Sy1327
MU10		3052	2858	3207	1716	1605	1531 1/83	
	4-C1	0002	2928	3291	1649	1359	1551,1405	v (C-CI)/01
MU11		2011	2884	3246	1721	1606	1510 1100	(∩⊔\ 2265
	4 - OH	3041	2931	3281	1664	1347	1342,1400	v (UH) 3303





Figure (11): The compound's FT-IR spectra (MH9).





In the 1H-NMR spectrum of MH10, two triple signals were observed attributed to (CH_2-CH_2) in the heptad ring in the range (2.90-3.24) ppm, and a signal at (8.65) ppm because of the presence of proton (CH) in the resulting ring, in addition to two signals in the range (6.15-6.42) ppm attributed to the double bond (CH=CH), and several signals noticed in the range (7.19-7.95) ppm attributed to the protons of the aromatic ring, and a signal at positions (6.80) ppm because of the presence of proton (NH) adjacent to the triazole ring, and two signals at positions (9.27-9.63) ppm because of the presence of proton (NH), and a signal at position (12.21) ppm because of the presence of proton (SH), then the solvent signal DMSO-d6 at 2.49 ppm, as shown in Figure 13.





Figure (13): 1-H NMR spectra of the substance (MH10).

In the 13C-NMR spectrum of MH10, two signals were observed attributed to the carbon of the (CH₂-CH₂) ring formed at positions (28.32, 34.48) ppm, a signal at (179.75) ppm because of the presence of carbonyl of the ester, a signal at 177.04 ppm attributed to the carbonyl of the amide, a signal at position (90.30) ppm because of the presence of (CH) the seven-membered ring, and the carbons of the aromatic ring appear in the range (120.11-142.63) ppm, and between the spectrum there are two signals at (11.91-119.74) ppm because of the presence of carbon of the (CH=CH) five-membered ring, and two signals at (164.08-159.95) ppm attributed to (C=N) the triazole ring. The protons of the solvent DMSO-d6 appear between 39.37 and 40.62 ppm, as in Figure 14.



Figure (14): ¹³C-NMR Spectra of the substance (MH10).



3.3. Evaluation of the Biological Activity of Prepared Compounds

The antibacterial activity of the prepared compounds was tested using the agar diffusion method. After inoculating the culture medium with the bacterial isolates[29,30], holes were made in the Petri dishes using the cylinder measuring method (according to USP 35). Using a drill: Place the prepared compounds (40 μ l) at three concentrations in each well and incubate the dish at (37°C) (24 hours) before taking the results. The readings were taken after (24) hours and (48) hours to indicate the derivatives' sensitivity. This depends on the apparent inhibitory diameter in the Petri dish surrounding the wells used; the increase in inhibitory diameter means an increase in the inhibitory diameter [31-33]. The bioavailability of the synthesised compounds was measured and compared with the inhibitory diameter of the standard antibiotics, some of which were used as control samples in the form of a solution[34-40].

Comp. No.	Klebse	illa pneu	moniae	e Staph. Epidermidis		
	0.001	0.01	0.1	0.001	0.01	0.1
MH1	0.2	0.2	1	0.1	0.2	1.1
MH2	0.2	0.5	0.8	0.2	0.2	0.8
MH3	0.3	0.8	1.2	1	1	1.5
NH5	0.2	0.5	2	0.5	1	1.5
MH8	0.5	0.5	0.8	0.5	1.2	2
MH9	0.2	0.5	1.2	1.5	1.9	2.2
MH11	0.2	0.8	1	0.5	3.5	4.5
MH12	0.4	0.8	2.3	0.8	1.6	2.2
Amoxicillin	21	22	25	25	29	33

Table (5): Biological efficiency of some synthesised compounds and control parameters (cm).



Figure (7): Biological effectiveness of the compound MH5, MH12 against bacterial K.pneumoniae.





Figure (8): Biological effectiveness of the compound MH9, MH11 against bacterial *Staph. epidermidis*.

1. Conclusions

The current study applies infrared and nuclear magnetic resonance spectra to explore the bacterial efficacy of some oxazepane derivatives derived from hydrazones.

The validity of the compositions of the synthesised compounds was confirmed by the physical qualities of the melting points and colours and by spectral studies, as well as the infrared spectrum and nuclear magnetic resonance spectrum of the proton and carbon. When testing its bacterial sensitivity against bacterial species, they gave good efficacy against Gram-negative and positive bacteria in comparison with antibiotics such as *Amoxicillin*.

For future studies, it is recommended to study the possibility of applying the same techniques to explore the derivatives from other chemicals.

Conflicts of Interest: The authors declare no conflict of interest.

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